

Identification of Phospholipid Species Implicated in Dementia by Untargeted High Resolution LC/MS and Data Dependent MS/MS

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INTRODUCTION

Alzheimer's disease (AD) is the leading cause of dementia in the elderly. Recently, Mapstone et al.¹ reported a panel of plasma phospholipids (Table 1) that predicted cognitively normal adults who later progressed to either mild cognitive impairment or dementia due to AD. This study utilized a targeted metabolomics p180 kit (Biocrates, Life Science AG, Austria) that measures phospholipids by infusion and selected reaction monitoring (SRM) with a triple quadrupole mass spectrometer. Since phospholipids have many isomers and isobars comprised of different fatty acid and alkyl/alkenyl ether combinations, it is not possible to assign unequivocally the phospholipid species using a low resolution approach.

Identification of fatty acid constituents in phospholipids implicated in AD is critical for two reasons: (a) to determine the functions of these lipids and their contributions to the pathophysiology of disease; and (b) to determine the molecular identities so one can develop quantitative assays to measure these particular phospholipids in human plasma.

The objective of this study is to determine which fatty acids comprise phosphatidylcholine (PC) lipids including lyso-PC 18:2; PC 36:6, 38:0, 38:6, 40:1, 40:2, 40:6 and 40:6e (ether) in human plasma using liquid chromatography and high-resolution accurate mass and tandem mass spectrometry (LC-MS/MS).

These results demonstrate that high resolution accurate mass (HRAM) LC-MS/MS is an excellent method for identifying isomeric lipid species from human plasma and which is essential for discovering the most meaningful changes between normal and diseased states.

MATERIALS AND METHODS

LC-MS Sample Preparation. Aliquots of human plasma (30 μ L) from 5 normal, 5 MCI (mild cognitive impairment) and 5 AD Alzheimer's-type dementia patients were extracted using the method of Bligh and Dyer. The organic phase was evaporated to dryness and the lipid extract was reconstituted in 100 μ L of 50:50 IPA/methanol. A pooled sample was prepared and injected in duplicate for positive and negative ion runs.

LC-MS and dd-MS² Method. A ThermoFisher Scientific™ Dionex™ UltiMate™ 3000 RSLC system and ThermoFisher Scientific™ Orbitrap Fusion Lumos™ mass spectrometer were employed. The HPLC separation (Table 2) was achieved with a 2.1 x 150mm, 3.0 μ m, Thermo Scientific C30 Acclaim™ column² and MS analysis (Table 3) was performed at 120K resolution (FWHM at m/z 200) and untargeted data dependent HCD MS² at 15K resolution. When a phosphocholine (m/z 184.0733) product ion was observed during the profiling experiment, a single CID experiment was also performed on the same precursor ion³.

DATA PROCESSING

Thermo Scientific™ LipidSearch 4.1™ software was used for lipid identification using the workflow in Figure 1. The LC-MS data (4 raw files, 2.3 GB total) containing high resolution MS and data dependent MS² were searched using the parameters in Table 4. For each MS² spectrum, results are obtained for those lipid species matching the accurate m/z of the precursor ion and its predicted fragment ions. Lipid annotations from all 4 files were correlated within ± 0.1 min and search results were merged into a single table. In this study, molecular lipid annotations were reported only if the ID's correlated in both positive and negative ion modes.

RESULTS

The LipidSearch results for PC 40:6 illustrates the identification process (Figure 2). Three different adducts were observed for the PC 40:6 species including M+H⁺ (m/z 834.6007), M+Na⁺ (m/z 856.5827) and M+HCO₂⁻ (m/z 878.5916). Mass chromatograms (Figure 2a and 2b) are displayed for formate and protonated adduct ions at retention time 15.1 min.

The negative ion HCD MS² spectrum (Figure 2c) matches PC containing 22:6 and 18:0 fatty acids: neutral loss (NL) of 18:0 and 22:6 ketene from the M-CH₃ anion (m/z 818.5705) are observed at m/z 552.3096 and 508.3409, respectively, in addition to the fatty acid anions. A choline containing head group in negative ion is inferred by NL of methyl formate.

The positive ion HCD and CID MS² spectra combined give the phosphocholine fragment ion (Figure 2d, m/z 184.0733) and NL of fatty acid / ketene: -22:6 (m/z 506.3605 / 524.3711); -18:0 (m/z 524.3711 / 552.3102) confirming the negative ion results. The underscore (18:0_22:6) is used to denote the fatty acyl position on the glycerol is not assigned in this case.

Similarly, all species in Table 1 were investigated to determine the fatty acid compositions and the identification results are summarized in Table 5. Although some of the previously identified sum composition species are present, there were instances of incorrect assignments, based on the SRM data, and mixtures of multiple species including false positives.

Figure 3 shows that three different PC 38:6 isomers are observed in the negative ion mass chromatogram of the formate adduct, m/z 850.5604. The MS² spectrum of isomer (1) matches the fragment ions predicted for 18:2_20:4; isomer (2), 18:1_20:5; and isomer (3), 16:0_22:6. NL of 20:4, 20:5 and 22:6 ketene from the M-CH₃ anions, and formation of a pair of fatty acid anions provide unambiguous annotations.

Table 1. Lipid Species Identified in Previous Studies That May Be Potential Indicators of Dementia Status

Lipid Class	Species (#C : # DB)	Elemental Composition	m/z [M+H] ⁺	m/z [M+HCO ₂] ⁻
LPC	18:2	C ₂₆ H ₅₀ N O ₇ P	520.3398	564.3306
PC	36:6	C ₄₄ H ₇₆ N O ₈ P	778.5381	822.5291
PC	38:6	C ₄₆ H ₈₀ N O ₈ P	806.5694	850.5604
PC	38:0	C ₄₆ H ₉₂ N O ₈ P	818.6633	862.6543
PC	40:6e	C ₄₈ H ₈₆ N O ₇ P	820.6215	864.6124
PC	40:6	C ₄₈ H ₈₄ N O ₈ P	834.6007	878.5954
PC	40:2	C ₄₈ H ₉₂ N O ₈ P	842.6633	886.6543
PC	40:1	C ₄₈ H ₉₄ N O ₈ P	844.6790	888.6700

Table 2. HPLC Method and Operating Conditions

Time, min	% A	% B	HPLC Parameters	Conditions
0	70	30	Mobile phase A	60:40 CH ₃ CN / H ₂ O
2	57	43	Mobile phase B	90:10 IPA / CH ₃ CN
2.1	45	55	Mobile phase B	90:10 IPA / CH ₃ CN
12	35	65	Mobile phase B	90:10 IPA / CH ₃ CN
18	15	85	buffer (A & B)	10mM NH ₄ HCO ₂ + 0.1% HCO ₂ H
20	0	100	Flow rate	260 μ L/min
25	0	100	Column Temp	45 ° C
25.1	70	30	Column Temp	45 ° C
30	70	30	Injection vol.	2 μ L

Table 3. High Resolution LC-MS Profiling Method

HESI Source	Orbitrap Fusion Lumos MS
Sheath Gas: 40	Pos. Ion (250–1200 amu); Neg. Ion (200–1200 amu)
Aux Gas: 3	MS Resolution: R = 120K (FWHM m/z 200)
Spray Voltage: 3.5 kV (2.5 kV neg. ion)	Top-Speed dd-MS ² : 1.0 sec at 15K (FWHM m/z 200)
RF-Lens: 50	MS ² Isolation Width: 1.0 Da
Cap. Temp: 320 C	Stepped NCE – Pos. 27 \pm 3; Neg. 30 \pm 10
Heater Temp: 300 C	AGC Target: 4.0 E+5 MS, 50 msec max. 5.0 E+4 MS ² , 35 msec max. for 1.0 sec HCD m/z 184.0733 triggered CID MS ² (30%)

Table 4. LipidSearch 4.1 SP1 Parameters

Parameter	Settings
Prec. mass tol.	5 ppm
Prod. mass tol.	5 ppm
Phospholipids	LPC, PC, LPE, PE, LPS, PS, LPG, PG, LPI, PI, LPA, PA, CL
Sphingolipids	So, LSM, SM, Cer, CerG1, CerG2, CerG3
Glycerolipids	MG, DG, TG
Neutral lipids	ChE, CoQ
Other	Acylcarnitines

Figure 1. LipidSearch Workflow

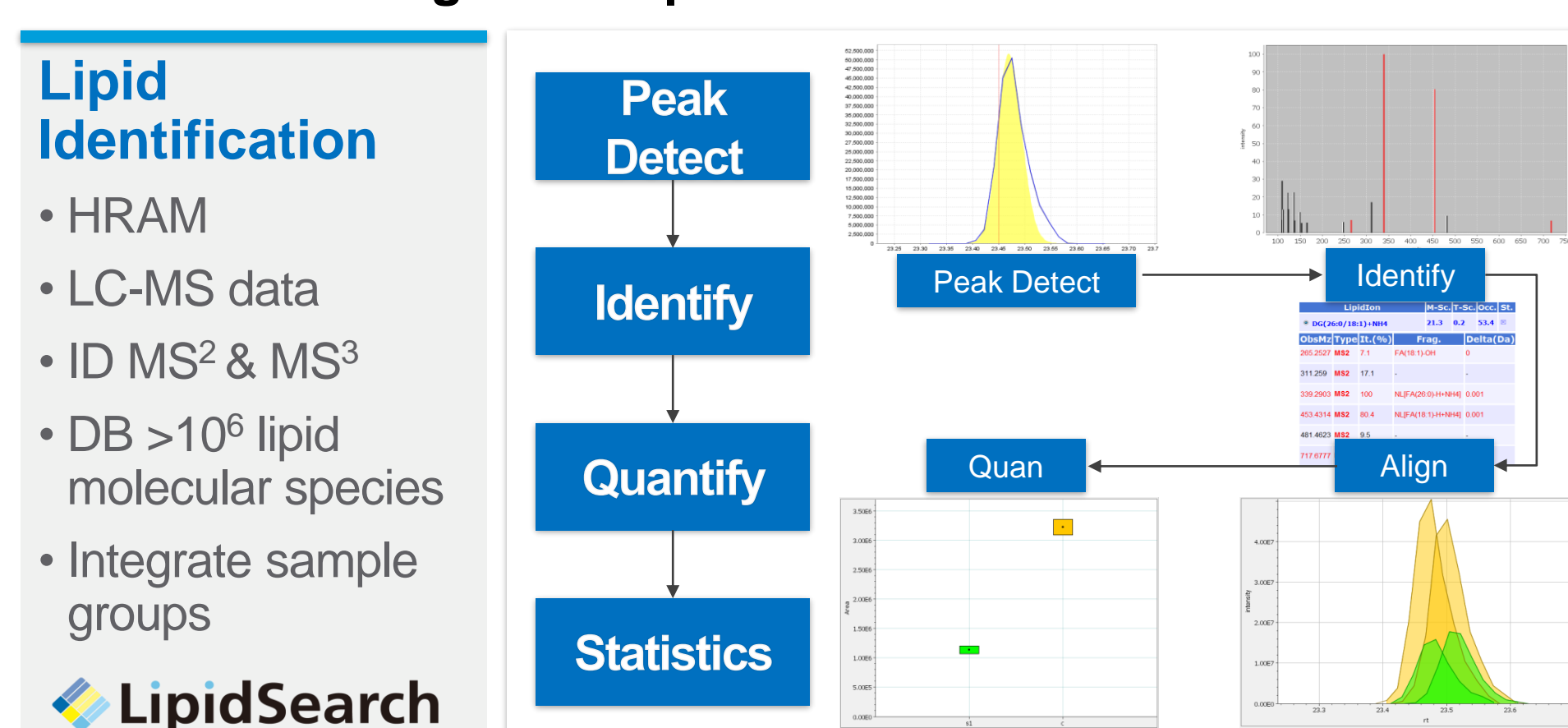


Figure 2. LipidSearch Identification of 18:0_22:6 PC

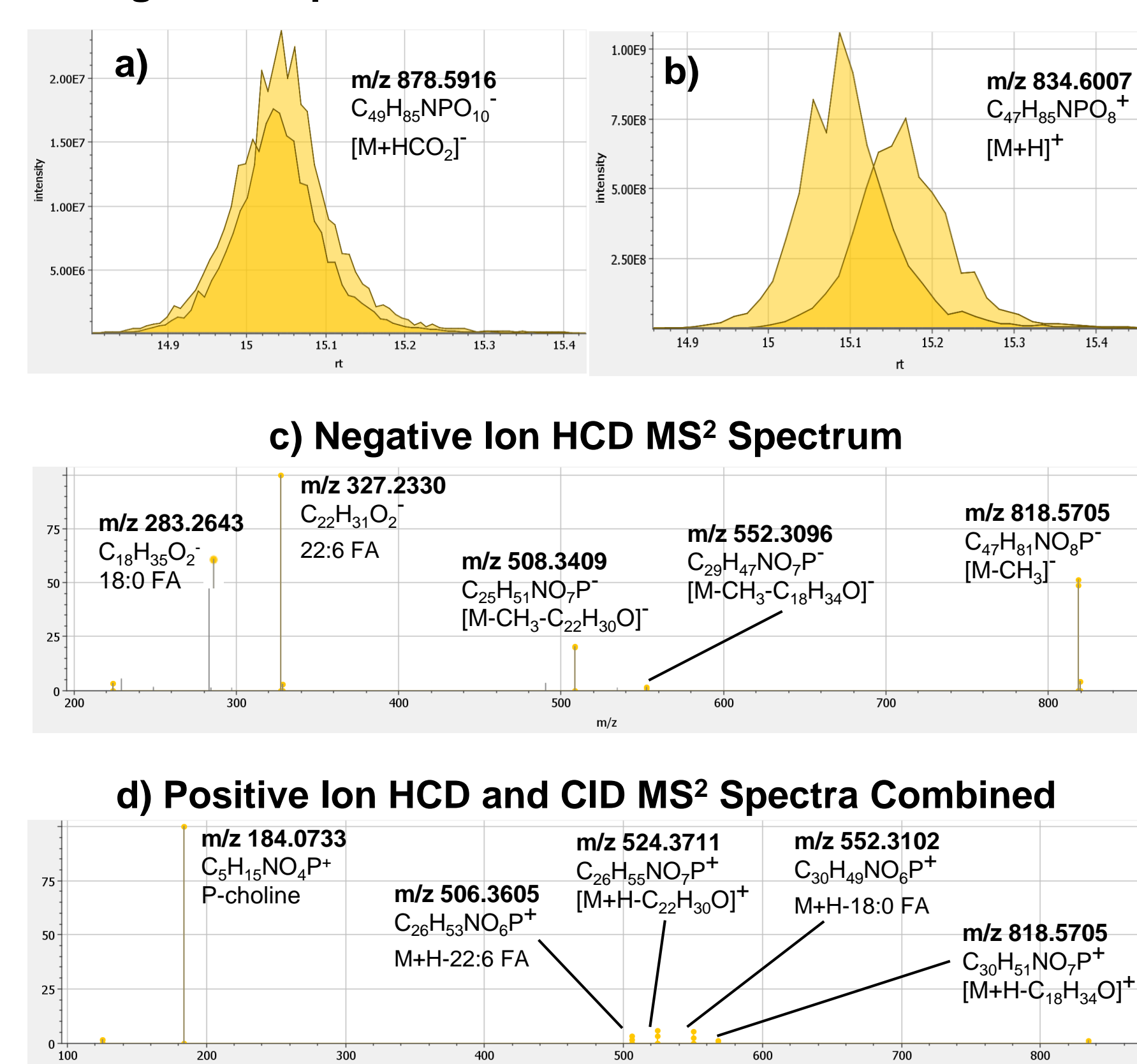
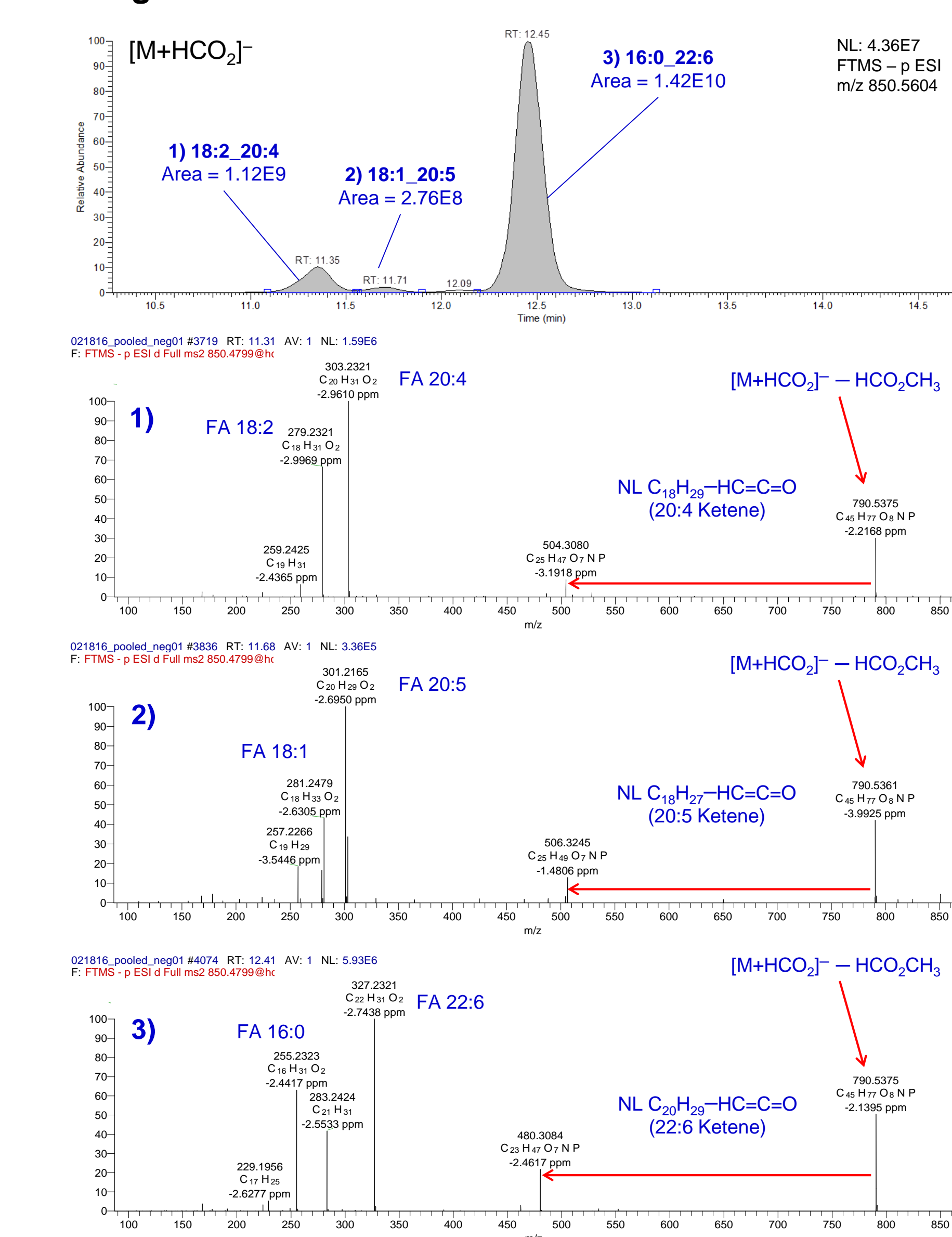


Table 5. Phosphatidylcholine Molecular Species Identified by High Resolution LC-MS and dd-MS²

Rt min	PC	Molecular Species	m/z [M+H] ⁺	m/z [M+HCO ₂] ⁻	Confirmed? - Comments
3.8	Lyso 18:2	Lyso 18:2	520.3398	564.3307	Yes; HCD - / CID +
10.2	36:6	14:0_22:6	778.5381	822.5291	Yes; HCD - / CID +
11.4	38:6	18:2_20:4	806.5694	850.5604	Yes; HCD - / CID +
11.7	38:6	18:1_20:5	806.5694	850.5604	Yes; HCD - / CID +
12.5	38:6	16:0_22:6	806.5694	850.5604	Yes; HCD - / CID +
19.5	38:0	38:0	818.6633	862.6543	No; HCD - mix; + m/z 184
15.9	40:7e	18:1E_22:6	818.6058	862.5967	Yes; HCD - / CID +
15.0	40:6e	18:1E_22:5	820.6215	864.6124	Yes; HCD -
16.3	40:6e	20:2E_20:4	820.6215	864.6124	Yes; HCD - / CID +
15.1	40:6	18:0_22:6	834.6007	878.5917	Yes; HCD - / CID +
18.9	40:2	22:0_18:2	842.6633	886.6543	Yes; HCD - / CID +
19.6	40:1	40:1	844.6790	888.6699	No; No MS ² data

Figure 3. Identification of Three Main 38:6 PC Isomers



DISCUSSION

PC molecular species confirmed in this study are Lyso 18:2, 36:6, 38:6, 40:2, 40:6 and 40:6e. Previously miss-identified 38:0 PC was found to be instead 40:7e; a low mass resolution SRM experiment (m/z 818.6 > 184.1) does not distinguish 38:0 from 40:7e PC. However, high resolution accurate mass measurements of precursor (m/z 818.6633 pos. ion, 862.5967 neg. ion) and product ions from LC-MS² data unequivocally assigns 40:7e as the species of interest. PC 40:1 was not confirmed due to extremely low abundance.

CONCLUSIONS

- Orbitrap-based LC-MS/MS untargeted lipidomics and LipidSearch software enables reliable and comprehensive lipid identification. This method was successfully applied to identification of lipid species implicated in AD patients.
- High mass accuracy and specificity of the LC-MS (120K) and MS² HCD/CID identification (15K) obtained with the Fusion Lumos MS allows confident lipid identification from human plasma extracts.
- Profiling of lipids in human plasma by flow injection SRM may lead to false positives or incorrect assignments.

REFERENCES

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